

OSMASEM and TSSEM

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```
# Data preparation
```

```
library(metaSEM)
```

```
## Loading required package: OpenMx
```

```
## To take full advantage of multiple cores, use:
```

```
##   mxOption(NULL, 'Number of Threads', parallel::detectCores()) #now
```

```
##   Sys.setenv(OMP_NUM_THREADS=parallel::detectCores()) #before library(OpenMx)
```

```
## "SLSQP" is set as the default optimizer in OpenMx.
```

```
## mxOption(NULL, "Gradient algorithm") is set at "central".
```

```
## mxOption(NULL, "Optimality tolerance") is set at "6.3e-14".
```

```
## mxOption(NULL, "Gradient iterations") is set at "2".
```

```
## Read List
```

```
ewomDataNew2.culture <- readRDS("ewomDataNew2.culture.RData")
```

```
## Check the number of studies per correlation coefficient
```

```
pattern.na(ewomDataNew2.culture$Data, show.na = FALSE)
```

```
## ReceiverInvolvement ResponseAdoptionWOM ResponsePH
```

```
## ReceiverInvolvement 62 6 5
```

```
## ResponseAdoptionWOM 6 62 23
```

```
## ResponsePH 5 23 62
```

```
## SourceCredibility 8 22 26
```

```
## WOMQualCred 9 27 32
```

```
## SourceCredibility WOMQualCred
```

```
## ReceiverInvolvement 8 9
```

```
## ResponseAdoptionWOM 22 27
```

```
## ResponsePH 26 32
```

```
## SourceCredibility 62 41
```

```
## WOMQualCred 41 62
```

```
## Calculate the sampling covariance matrix of the correlations
```

```
my.df_ewom <- Cor2DataFrame(ewomDataNew2.culture$Data, ewomDataNew2.culture$n, acov = "weighted")
```

```
# One-Stage MASEM
```

```
## Model without moderation
```

```
ewom_model1 <- 'ResponseAdoptionWOM ~ PH2Adopt*ResponsePH
```

```
ResponsePH ~ INV2PH*ReceiverInvolvement + Qual2PH*WOMQualCred + SC2PH*SourceCredibility
```

```
WOMQualCred ~ SC2Qual*SourceCredibility
```

```
SourceCredibility ~~ scWITHinv*ReceiverInvolvement
```

```
SourceCredibility ~~ 1*SourceCredibility
```

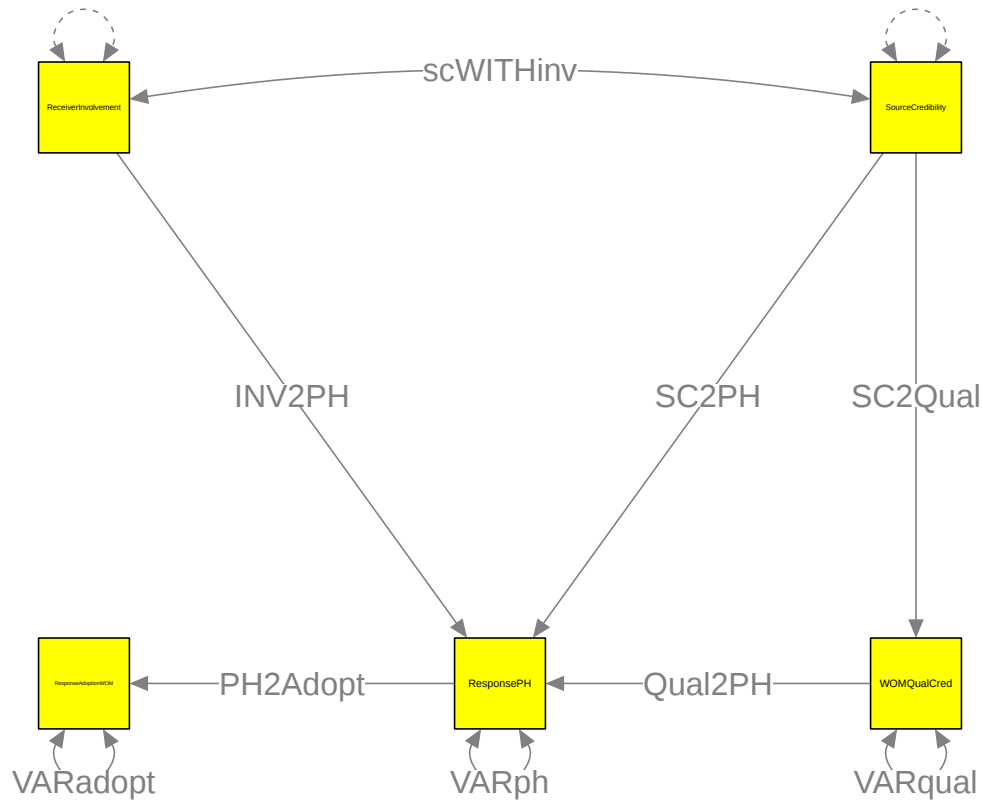
```
ReceiverInvolvement ~~ 1*ReceiverInvolvement
```

```
WOMQualCred ~~ VARqual*WOMQualCred
```

```
ResponsePH ~~ VARph*ResponsePH
```

```
ResponseAdoptionWOM ~~ VARadopt*ResponseAdoptionWOM'
```

```
plot(ewom_model1, col="yellow")
```



```
## Convert the lavaan syntax into the RAM specification
ewom_RAM1 <- lavaan2RAM(ewom_model1, obs.variables=c("ReceiverInvolvement", "ResponseAdoptionWOM",
                                                    "ResponsePH", "SourceCredibility", "WOMQualCred"))
ewom_RAM1
```

```
## $A
##           ReceiverInvolvement ResponseAdoptionWOM ResponsePH
## ReceiverInvolvement "0"           "0"           "0"
## ResponseAdoptionWOM "0"           "0"           "0*PH2Adopt"
## ResponsePH          "0*INV2PH"    "0"           "0"
## SourceCredibility   "0"           "0"           "0"
## WOMQualCred         "0"           "0"           "0"
##
##           SourceCredibility WOMQualCred
## ReceiverInvolvement "0"       "0"
## ResponseAdoptionWOM "0"       "0"
## ResponsePH          "0*SC2PH" "0*Qual2PH"
## SourceCredibility   "0"       "0"
## WOMQualCred         "0*SC2Qual" "0"
##
## $S
##           ReceiverInvolvement ResponseAdoptionWOM ResponsePH
## ReceiverInvolvement "1"       "0"           "0"
## ResponseAdoptionWOM "0"       "0*VARadopt"    "0"
## ResponsePH          "0"       "0"           "0*VARph"
## SourceCredibility   "0*scWITHinv" "0"       "0"
## WOMQualCred         "0"       "0"           "0"
```

```

## SourceCredibility WOMQualCred
## ReceiverInvolvement "0*scWITHinv" "0"
## ResponseAdoptionWOM "0" "0"
## ResponsePH "0" "0"
## SourceCredibility "1" "0"
## WOMQualCred "0" "0*VARqual"
##
## $F
## ReceiverInvolvement ResponseAdoptionWOM ResponsePH
## ReceiverInvolvement 1 0 0
## ResponseAdoptionWOM 0 1 0
## ResponsePH 0 0 1
## SourceCredibility 0 0 0
## WOMQualCred 0 0 0
## SourceCredibility WOMQualCred
## ReceiverInvolvement 0 0
## ResponseAdoptionWOM 0 0
## ResponsePH 0 0
## SourceCredibility 1 0
## WOMQualCred 0 1
##
## $M
## ReceiverInvolvement ResponseAdoptionWOM ResponsePH SourceCredibility
## 1 0 0 0 0
## WOMQualCred
## 1 0
## Model without moderator
## Create the model implied correlation structure with implicit diagonal constraints
M0 <- create.vechsR(A0=ewom_RAM1$A, S0=ewom_RAM1$S)

## Create the heterogeneity variance-covariance matrix
## RE.type= either "Diag" or "Symm"
## Transform= either "expLog" or "sqSD" for better estimation of variances
T0 <- create.Tau2(RAM=ewom_RAM1, RE.type="Diag", Transform="expLog", RE.startvalues=0.05)

## Fit the model using One-Stage MASEM
mx.fit0_ewom <- osmasem(model.name="eWoM No moderator", Mmatrix=M0, Tmatrix=T0, data=my.df_ewom)
summary(mx.fit0_ewom, Saturated = TRUE)

## Summary of eWoM No moderator
##
## free parameters:
## name matrix row col Estimate
## 1 INV2PH A0 ResponsePH ReceiverInvolvement 0.3918654
## 2 PH2Adopt A0 ResponseAdoptionWOM ResponsePH 0.8123080
## 3 SC2PH A0 ResponsePH SourceCredibility 0.2180440
## 4 SC2Qual A0 WOMQualCred SourceCredibility 0.5354310
## 5 Qual2PH A0 ResponsePH WOMQualCred 0.4461319
## 6 scWITHinv S0 SourceCredibility ReceiverInvolvement 0.2687476
## 7 Tau1_1 vecTau1 1 1 -1.8847894
## 8 Tau1_2 vecTau1 2 1 -1.8572631
## 9 Tau1_3 vecTau1 3 1 -2.3683576
## 10 Tau1_4 vecTau1 4 1 -1.2437278
## 11 Tau1_5 vecTau1 5 1 -1.2300574

```

```

## 12   Tau1_6 vecTau1          6          1 -1.9897811
## 13   Tau1_7 vecTau1          7          1 -2.0306503
## 14   Tau1_8 vecTau1          8          1 -2.0766360
## 15   Tau1_9 vecTau1          9          1 -1.7814900
## 16   Tau1_10 vecTau1         10          1 -1.7717364
##      Std.Error A      z value      Pr(>|z|)
## 1  0.06270486    6.249362 4.121328e-10
## 2  0.04827675   16.826071 0.000000e+00
## 3  0.04468874    4.879171 1.065329e-06
## 4  0.02804820   19.089672 0.000000e+00
## 5  0.04181465   10.669272 0.000000e+00
## 6  0.04017397    6.689595 2.237899e-11
## 7  0.32661353   -5.770702 7.894182e-09
## 8  0.34115406   -5.444060 5.207975e-08
## 9  0.36234408   -6.536212 6.309664e-11
## 10 0.25250535   -4.925550 8.412332e-07
## 11 0.20375813   -6.036850 1.571513e-09
## 12 0.16812472  -11.835149 0.000000e+00
## 13 0.16214650  -12.523553 0.000000e+00
## 14 0.15870144  -13.085174 0.000000e+00
## 15 0.14250529  -12.501220 0.000000e+00
## 16 0.11616287  -15.252175 0.000000e+00
##
## Model Statistics:
##      | Parameters | Degrees of Freedom | Fit (-2lnL units)
##      Model:          16          183          -137.1828
##      Saturated:       20          179          -185.5389
##      Independence:    20          179           NA
## Number of observations/statistics: 28516/199
##
## chi-square:   $\chi^2$  ( df=4 ) = 48.35611,  p = 7.954756e-10
## Information Criteria:
##      | df Penalty | Parameters Penalty | Sample-Size Adjusted
##      AIC:      -503.1828          -105.18281          -105.16373
##      BIC:      -2014.4372           26.94872           -23.89902
##      CFI: NA
##      TLI: NA (also known as NNFI)
##      RMSEA: 0.0197198 [95% CI (0.01401375, 0.02582225)]
##      Prob(RMSEA <= 0.05): 1
##      To get additional fit indices, see help(mxRefModels)
##      timestamp: 2019-04-24 12:53:32
##      Wall clock time: 1.460166 secs
##      optimizer: SLSQP
##      OpenMx version number: 2.12.2
##      Need help? See help(mxSummary)
osmasemSRMR(mx.fit0_ewom)

## [1] 0.1150122
## The variance-covariance matrix in mx.fit0 is based on the untransformed matrix
## Extract the heterogeneity variance-covariance matrix using VarCorr()
VarCorr(mx.fit0_ewom)

##      Tau2_1      Tau2_2      Tau2_3      Tau2_4      Tau2_5      Tau2_6

```

```
## Tau2_1 0.02306177 0.00000000 0.00000000 0.00000000 0.00000000 0.00000000
## Tau2_2 0.00000000 0.02436699 0.00000000 0.00000000 0.00000000 0.00000000
## Tau2_3 0.00000000 0.00000000 0.008767399 0.00000000 0.00000000 0.00000000
## Tau2_4 0.00000000 0.00000000 0.00000000 0.08312119 0.00000000 0.00000000
## Tau2_5 0.00000000 0.00000000 0.00000000 0.00000000 0.08542515 0.00000000
## Tau2_6 0.00000000 0.00000000 0.00000000 0.00000000 0.00000000 0.01869382
## Tau2_7 0.00000000 0.00000000 0.00000000 0.00000000 0.00000000 0.00000000
## Tau2_8 0.00000000 0.00000000 0.00000000 0.00000000 0.00000000 0.00000000
## Tau2_9 0.00000000 0.00000000 0.00000000 0.00000000 0.00000000 0.00000000
## Tau2_10 0.00000000 0.00000000 0.00000000 0.00000000 0.00000000 0.00000000
##          Tau2_7      Tau2_8      Tau2_9      Tau2_10
## Tau2_1 0.00000000 0.00000000 0.00000000 0.00000000
## Tau2_2 0.00000000 0.00000000 0.00000000 0.00000000
## Tau2_3 0.00000000 0.00000000 0.00000000 0.00000000
## Tau2_4 0.00000000 0.00000000 0.00000000 0.00000000
## Tau2_5 0.00000000 0.00000000 0.00000000 0.00000000
## Tau2_6 0.00000000 0.00000000 0.00000000 0.00000000
## Tau2_7 0.0172266 0.00000000 0.00000000 0.00000000
## Tau2_8 0.00000000 0.01571292 0.00000000 0.00000000
## Tau2_9 0.00000000 0.00000000 0.02835421 0.00000000
## Tau2_10 0.00000000 0.00000000 0.00000000 0.02891274
```

```
# TSSEM
```

```
## Model without moderator
```

```
## Stage 1 analysis
```

```
ewom_random1 <- tssem1(ewomDataNew2.culture$Data, ewomDataNew2.culture$n, method="REM", RE.type="Diag")
summary(ewom_random1)
```

```
##
```

```
## Call:
```

```
## meta(y = ES, v = acovR, RE.constraints = Diag(paste0(RE.startvalues,
##      "*Tau2_", 1:no.es, "_", 1:no.es)), RE.lbound = RE.lbound,
##      I2 = I2, model.name = model.name, suppressWarnings = TRUE,
##      silent = silent, run = run)
```

```
##
```

```
## 95% confidence intervals: z statistic approximation
```

```
## Coefficients:
```

##	Estimate	Std.Error	lbound	ubound	z value	Pr(> z)
## Intercept1	0.4468965	0.0656584	0.3182083	0.5755847	6.8064	1.001e-11
## Intercept2	0.4936833	0.0734816	0.3496619	0.6377046	6.7185	1.837e-11
## Intercept3	0.2505928	0.0392090	0.1737446	0.3274411	6.3912	1.646e-10
## Intercept4	0.3988718	0.0480125	0.3047690	0.4929746	8.3077	< 2.2e-16
## Intercept5	0.5667571	0.0355506	0.4970792	0.6364350	15.9423	< 2.2e-16
## Intercept6	0.4886148	0.0312245	0.4274160	0.5498137	15.6485	< 2.2e-16
## Intercept7	0.5497398	0.0265882	0.4976279	0.6018518	20.6761	< 2.2e-16
## Intercept8	0.5405100	0.0270608	0.4874718	0.5935482	19.9739	< 2.2e-16
## Intercept9	0.5648842	0.0303301	0.5054383	0.6243302	18.6245	< 2.2e-16
## Intercept10	0.5306669	0.0284074	0.4749895	0.5863443	18.6806	< 2.2e-16
## Tau2_1_1	0.0212558	0.0149490	-0.0080437	0.0505554	1.4219	0.1550590
## Tau2_2_2	0.0235264	0.0167537	-0.0093102	0.0563631	1.4043	0.1602436
## Tau2_3_3	0.0085736	0.0062289	-0.0036348	0.0207821	1.3764	0.1686912
## Tau2_4_4	0.0168496	0.0102027	-0.0031474	0.0368466	1.6515	0.0986401
## Tau2_5_5	0.0252136	0.0082472	0.0090495	0.0413777	3.0572	0.0022338
## Tau2_6_6	0.0176311	0.0059478	0.0059737	0.0292886	2.9643	0.0030336

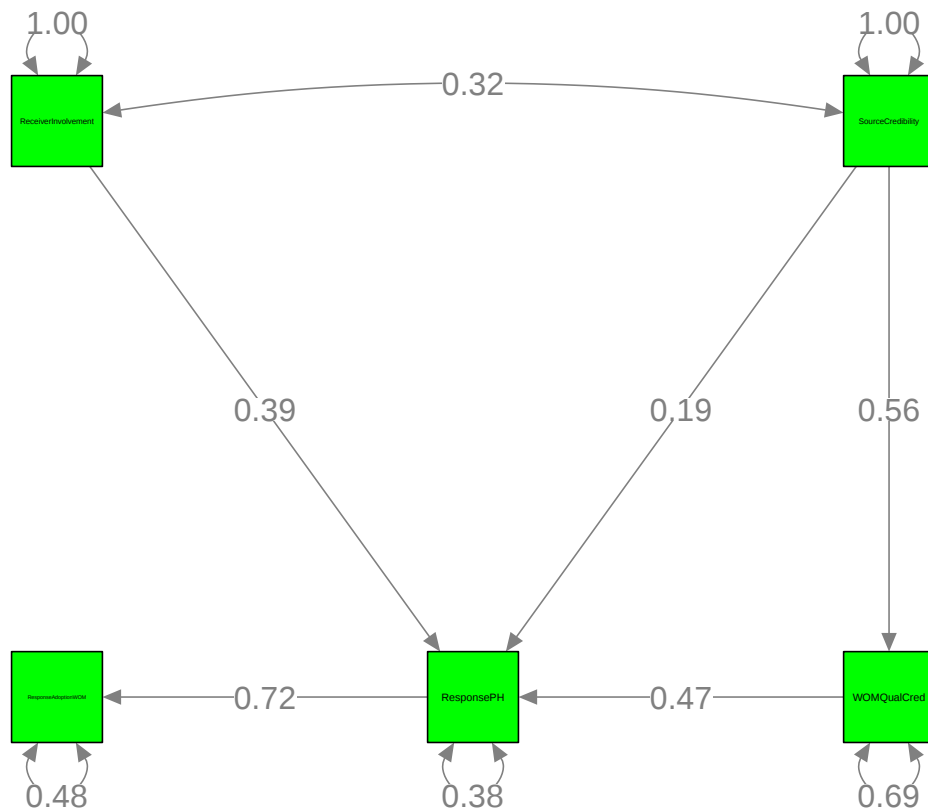
```

## Tau2_7_7      0.0152332  0.0046654  0.0060891  0.0243772  3.2651 0.0010941
## Tau2_8_8      0.0151144  0.0048948  0.0055208  0.0247079  3.0879 0.0020160
## Tau2_9_9      0.0255151  0.0069029  0.0119858  0.0390445  3.6963 0.0002188
## Tau2_10_10    0.0295540  0.0069100  0.0160106  0.0430975  4.2770 1.895e-05
##
## Intercept1    ***
## Intercept2    ***
## Intercept3    ***
## Intercept4    ***
## Intercept5    ***
## Intercept6    ***
## Intercept7    ***
## Intercept8    ***
## Intercept9    ***
## Intercept10   ***
## Tau2_1_1
## Tau2_2_2
## Tau2_3_3
## Tau2_4_4      .
## Tau2_5_5      **
## Tau2_6_6      **
## Tau2_7_7      **
## Tau2_8_8      **
## Tau2_9_9      ***
## Tau2_10_10    ***
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Q statistic on the homogeneity of effect sizes: 3761.543
## Degrees of freedom of the Q statistic: 189
## P value of the Q statistic: 0
##
## Heterogeneity indices (based on the estimated Tau2):
##
##               Estimate
## Intercept1: I2 (Q statistic)  0.9038
## Intercept2: I2 (Q statistic)  0.9123
## Intercept3: I2 (Q statistic)  0.7912
## Intercept4: I2 (Q statistic)  0.8818
## Intercept5: I2 (Q statistic)  0.9217
## Intercept6: I2 (Q statistic)  0.8890
## Intercept7: I2 (Q statistic)  0.8761
## Intercept8: I2 (Q statistic)  0.8766
## Intercept9: I2 (Q statistic)  0.9236
## Intercept10: I2 (Q statistic) 0.9372
##
## Number of studies (or clusters): 62
## Number of observed statistics: 199
## Number of estimated parameters: 20
## Degrees of freedom: 179
## -2 log likelihood: -179.5602
## OpenMx status1: 0 ("0" or "1": The optimization is considered fine.
## Other values may indicate problems.)

```

```
## Stage 2 analysis
ewom_random2 <- tssem2(ewom_random1, Amatrix=ewom_RAM1$A, Smatrix=ewom_RAM1$S,
                      Fmatrix = ewom_RAM1$F, diag.constraints = FALSE,
                      intervals.type = "z")
summary(ewom_random2)

##
## Call:
## wls(Cov = pooledS, aCov = aCov, n = tssem1.obj$total.n, Amatrix = Amatrix,
##      Smatrix = Smatrix, Fmatrix = Fmatrix, diag.constraints = diag.constraints,
##      cor.analysis = cor.analysis, intervals.type = intervals.type,
##      mx.algebras = mx.algebras, model.name = model.name, suppressWarnings = suppressWarnings,
##      silent = silent, run = run)
##
## 95% confidence intervals: z statistic approximation
## Coefficients:
##           Estimate Std. Error   lbound   ubound z value Pr(>|z|)
## PH2Adopt  0.722524  0.025698 0.672157 0.772892 28.1159 < 2.2e-16 ***
## INV2PH    0.390104  0.065992 0.260761 0.519446  5.9114 3.393e-09 ***
## SC2PH     0.193565  0.047918 0.099647 0.287484  4.0395 5.357e-05 ***
## Qual2PH   0.467647  0.038891 0.391421 0.543872 12.0245 < 2.2e-16 ***
## SC2Qual   0.556167  0.027911 0.501462 0.610872 19.9262 < 2.2e-16 ***
## scWITHinv 0.324695  0.036306 0.253536 0.395853  8.9432 < 2.2e-16 ***
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Goodness-of-fit indices:
##
##                                     Value
## Sample size                        28516.0000
## Chi-square of target model          70.2071
## DF of target model                   4.0000
## p value of target model              0.0000
## Number of constraints imposed on "Smatrix" 0.0000
## DF manually adjusted                 0.0000
## Chi-square of independence model     2073.9395
## DF of independence model             10.0000
## RMSEA                               0.0241
## RMSEA lower 95% CI                   0.0193
## RMSEA upper 95% CI                   0.0292
## SRMR                                0.1016
## TLI                                  0.9198
## CFI                                  0.9679
## AIC                                  62.2071
## BIC                                  29.1742
## OpenMx status1: 0 ("0" or "1": The optimization is considered fine.
## Other values indicate problems.)
## Plot the model
plot(ewom_random2, col="green")
```



```
osmasem_coef <- coef(mx.fit0_ewom)
```

```
tssem_coef <- coef(ewom_random2)
```

```
## OSMASEM coefficients
```

```
osmasem_coef[sort(names(osmasem_coef))]
```

```
## INV2PH PH2Adopt Qual2PH SC2PH SC2Qual scWITHinv
## 0.3918654 0.8123080 0.4461319 0.2180440 0.5354310 0.2687476
```

```
## TSSEM coefficients
```

```
tssem_coef[sort(names(tssem_coef))]
```

```
## INV2PH PH2Adopt Qual2PH SC2PH SC2Qual scWITHinv
## 0.3901037 0.7225243 0.4676467 0.1935655 0.5561668 0.3246947
```

```
## OSMASEM variance components
```

```
diag(VarCorr(mx.fit0_ewom))
```

```
## Tau2_1 Tau2_2 Tau2_3 Tau2_4 Tau2_5 Tau2_6
## 0.023061773 0.024366986 0.008767399 0.083121192 0.085425149 0.018693820
## Tau2_7 Tau2_8 Tau2_9 Tau2_10
## 0.017226600 0.015712919 0.028354206 0.028912744
```

```
## TSSEM variance components
```

```
coef(ewom_random1, select="random")
```

```
## Tau2_1_1 Tau2_2_2 Tau2_3_3 Tau2_4_4 Tau2_5_5 Tau2_6_6
## 0.021255837 0.023526429 0.008573639 0.016849620 0.025213596 0.017631114
## Tau2_7_7 Tau2_8_8 Tau2_9_9 Tau2_10_10
## 0.015233156 0.015114385 0.025515120 0.029554033
```

```

## Fit indices for OSMASEM
# chi-square:  chisq ( df=4 ) = 48.3561,  p = 7.954771e-10
# RMSEA:  0.01971979  [95% CI (0.01401375, 0.02582224)]
# SRMR:  0.1150122

## Fit indices for TSSEM
# chi-square:  chisq ( df=4 ) = 70.2071,  p = .0000
# RMSEA:  0.0241  [95% CI (0.0193, 0.0292)]
# SRMR:  0.1016

sessionInfo()

## R version 3.5.1 (2018-07-02)
## Platform: x86_64-pc-linux-gnu (64-bit)
## Running under: Ubuntu 18.04.2 LTS
##
## Matrix products: default
## BLAS: /opt/microsoft/roper/3.5.1/lib64/R/lib/libRblas.so
## LAPACK: /opt/microsoft/roper/3.5.1/lib64/R/lib/libRlapack.so
##
## locale:
##  [1] LC_CTYPE=en_US.UTF-8      LC_NUMERIC=C
##  [3] LC_TIME=en_US.UTF-8      LC_COLLATE=en_US.UTF-8
##  [5] LC_MONETARY=en_US.UTF-8  LC_MESSAGES=en_US.UTF-8
##  [7] LC_PAPER=en_US.UTF-8     LC_NAME=C
##  [9] LC_ADDRESS=C             LC_TELEPHONE=C
## [11] LC_MEASUREMENT=en_US.UTF-8 LC_IDENTIFICATION=C
##
## attached base packages:
## [1] stats      graphics  grDevices  utils      datasets  methods   base
##
## other attached packages:
## [1] metaSEM_1.2.1.1      OpenMx_2.12.2      RevoUtils_11.0.1
## [4] RevoUtilsMath_11.0.0
##
## loaded via a namespace (and not attached):
##  [1] nlme_3.1-137      RColorBrewer_1.1-2  mi_1.0
##  [4] tools_3.5.1      backports_1.1.3     R6_2.4.0
##  [7] d3Network_0.5.2.1  rpart_4.1-13        Hmisc_4.2-0
## [10] lazyeval_0.2.1    colorspace_1.4-0    nnet_7.3-12
## [13] tidyselect_0.2.5  gridExtra_2.3        mnormt_1.5-5
## [16] compiler_3.5.1    qgraph_1.6.1        fdrtool_1.2.15
## [19] htmlTable_1.13.1  scales_1.0.0        checkmate_1.9.1
## [22] mvtnorm_1.0-9     psych_1.8.12        pbapply_1.4-0
## [25] sem_3.1-9         stringr_1.4.0       digest_0.6.18
## [28] pbivnorm_0.6.0    foreign_0.8-71      minqa_1.2.4
## [31] rmarkdown_1.11    base64enc_0.1-3     jpeg_0.1-8
## [34] pkgconfig_2.0.2   htmltools_0.3.6     lme4_1.1-20
## [37] lisrelToR_0.1.4   htmlwidgets_1.3     rlang_0.3.1
## [40] rstudioapi_0.9.0  huge_1.3.0          gtools_3.8.1
## [43] acepack_1.4.1     dplyr_0.8.0.1       zip_2.0.0
## [46] magrittr_1.5      Formula_1.2-3       Matrix_1.2-15
## [49] Rcpp_1.0.0        munsell_0.5.0       abind_1.4-5
## [52] rockchalk_1.8.140 whisker_0.3-2       stringi_1.3.1

```

## [55] yaml_2.2.0	carData_3.0-2	MASS_7.3-51.1
## [58] plyr_1.8.4	matrixcalc_1.0-3	lavaan_0.6-3
## [61] grid_3.5.1	parallel_3.5.1	crayon_1.3.4
## [64] lattice_0.20-38	semPlot_1.1	kutils_1.64
## [67] splines_3.5.1	knitr_1.21	pillar_1.3.1
## [70] igraph_1.2.4	rjson_0.2.20	boot_1.3-20
## [73] corpcor_1.6.9	BDgraph_2.55	reshape2_1.4.3
## [76] stats4_3.5.1	XML_3.98-1.17	glue_1.3.0
## [79] evaluate_0.13	latticeExtra_0.6-28	data.table_1.12.0
## [82] png_0.1-7	nloptr_1.2.1	gtable_0.2.0
## [85] purrr_0.3.0	assertthat_0.2.0	ggplot2_3.1.0
## [88] xfun_0.5	openxlsx_4.1.0	xtable_1.8-3
## [91] semTools_0.5-1	coda_0.19-2	survival_2.43-3
## [94] glasso_1.10	tibble_2.0.1	arm_1.10-1
## [97] ggm_2.3	ellipse_0.4.1	cluster_2.0.7-1